

## Recombinant Factor IX fused with albumin (rFIX-FP) is underassigned by one-stage methods with silica as contact activator.

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### INTRODUCTION

One-stage (OS) methods show in general considerably lower assigned FIX potency for Factor IX fused with albumin, (albutrepenonakog alfa, rFIX-FP) than chromogenic substrate (CS) methods. The cause(s) of the discrepancy is not known.

*Aim:* Investigation of causes of discrepancy on rFIX-FP potency assignment by commercial and new variant OS methods and a CS method.

### MATERIALS AND METHODS

FIX sources: rFIX-FP (CSL Behring, Germany), plasma derived (pd) FIX concentrate (BPL, UK) and purified human FIX (Enzyme Research, USA), denoted pure pdFIX. The 4<sup>th</sup> and 5<sup>th</sup> IS FIX Concentrate (NIBSC, UK) were used as calibrators. Colloidal Silica Bindzil 309/220 (AkzoNobel, Sweden).

OS methods using A) APTT reagents Pathromtin SL (Siemens, Germany) and SynthAFax (IL, USA); B) purified phospholipids (PL) Phospholipid-TGT (Rossix, Mölndal, Sweden) and a platelet membrane like PL composition PL-PF3 (in-house), both without any contact activator and instead including FXIa with CaCl<sub>2</sub>; C) APTT reagents comprising Phospholipid-TGT (PL-TGT) and PL-PF3 with 1.8 µL colloidal silica (CSi) / mL PL emulsion. CS method: Rox Factor IX (Rossix) applied on ACL TOP 500 and manually in microplates.

### RESULTS

Fig. 1 shows close to 80% higher assigned FIX potencies for rFIX-FP with Rox Factor IX and SynthAFax as compared to Pathromtin SL. No difference was obtained for pdFIX.

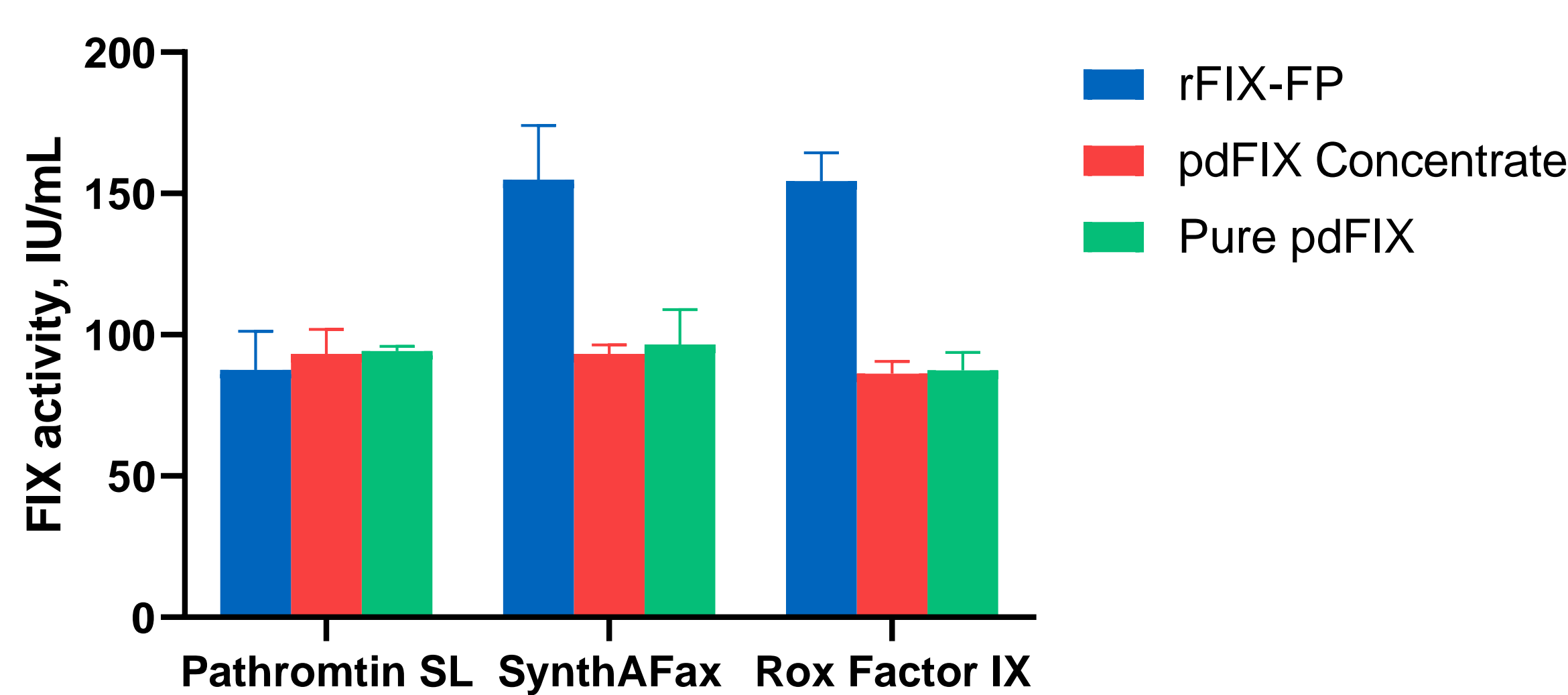
Fig. 2 shows that OS methods with PL-TGT and PL-PF3 and including FXIa as replacement of contact activation yield rFIX-FP potency similar to Rox Factor IX and SynthAFax. Addition of colloidal silica to both phospholipids drastically reduces rFIX-FP potency, yielding results similar to Pathromtin SL.

Fig. 3 shows a gradual decrease of rFIX-FP potency with increasing concentration of colloidal silica in the phospholipid emulsion.

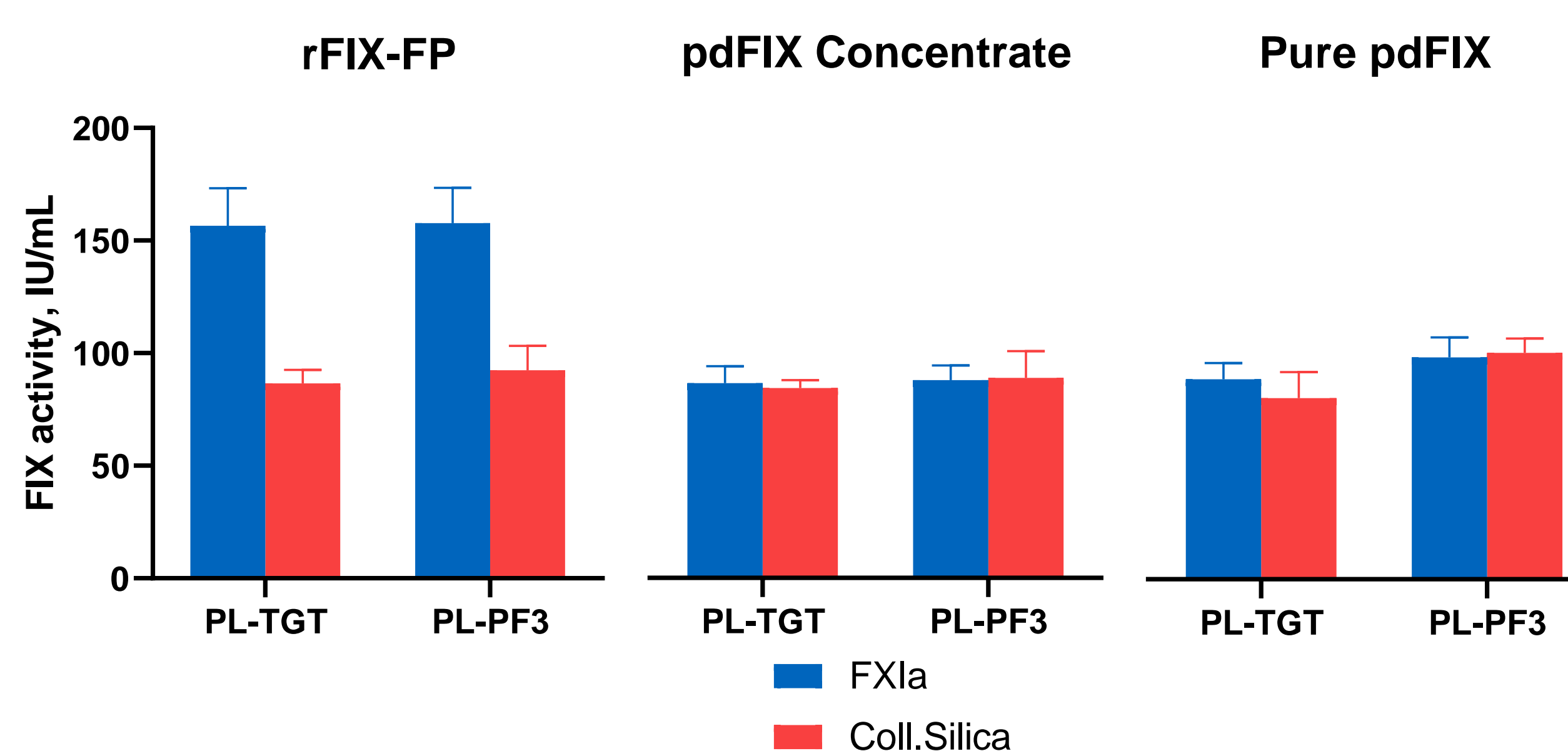
No anomaly was observed with Rox Factor IX for the activation kinetics of rFIX-FP vs the 5<sup>th</sup> IS (data not shown).

### CONCLUSIONS

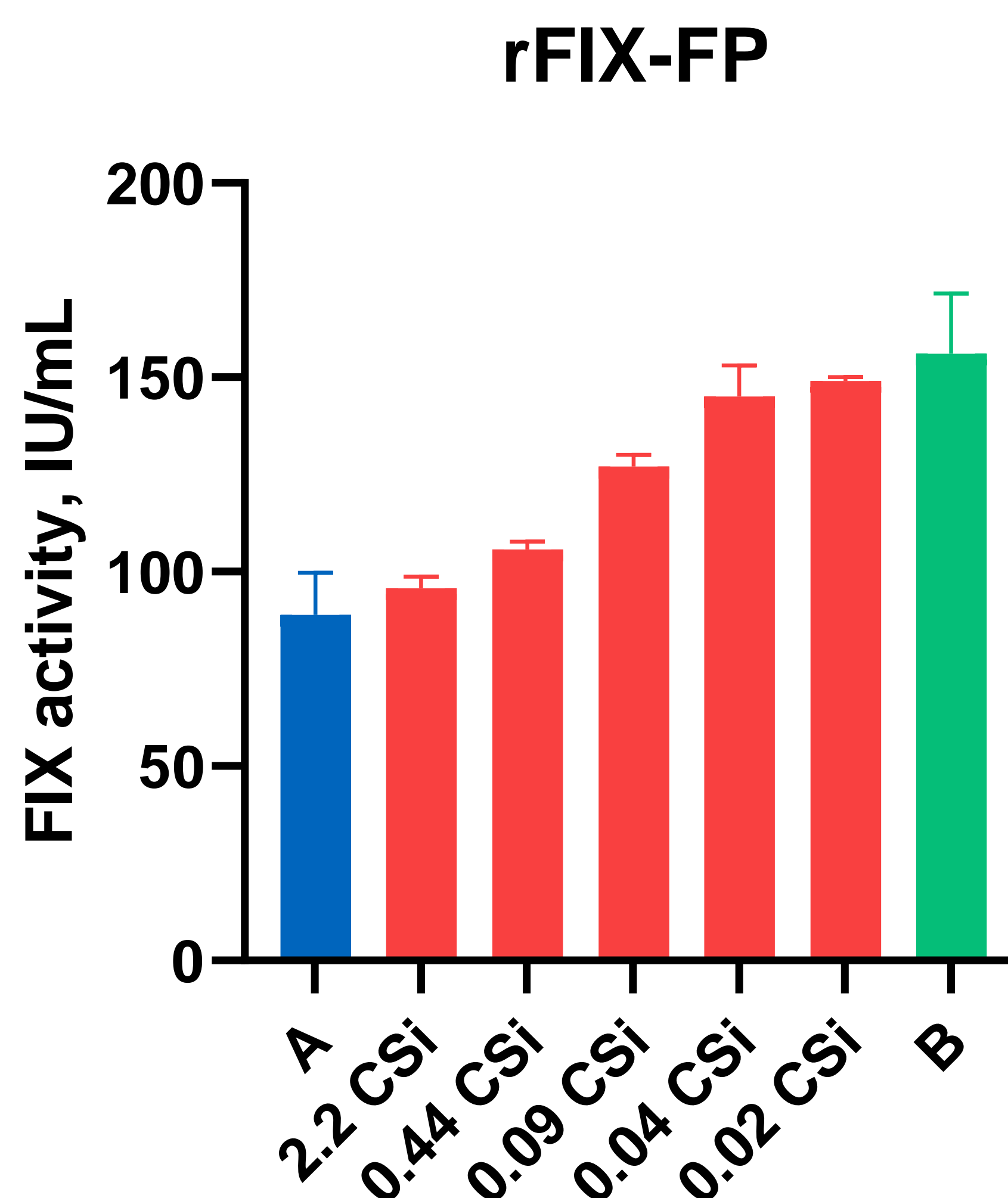
1. rFIX-FP potency is underassigned in OS methods using APTT reagents with colloidal silica as contact activator, most likely due to attenuated activation and hence deviation from like-vs-like behavior compared to a pdFIX concentrate standard.
2. OS methods with Phospholipid-TGT and a platelet membrane like PL composition, both with FXIa included with CaCl<sub>2</sub> (no contact activation), give results in line with Rox Factor IX and SynthAFax.
3. pdFIX potency is similar with all OS methods and with Rox Factor IX, thus displaying like-vs-like behavior to a pdFIX concentrate standard.



**Fig. 1** Mean FIX potencies obtained with Pathromtin SL, SynthAFax and Rox Factor IX. At least two dilutions were assayed of each sample with each method. rFIX-FP (n = 6-7), pdFIX Conc (n = 3-7), pure pdFIX (n = 3-4).



**Fig. 2** Mean FIX potencies obtained with OS methods using two phospholipid emulsions, PL-TGT and PL-PF3, either with added colloidal silica (1.8 µL/mL) or with FXIa (1.5 - 6 IU/mL) included in the calcium chloride solution, thus omitting contact activation.



**Fig. 3** Assigned mean FIX potencies of rFIX-FP in OS method with PL-TGT and with a range of colloidal silica concentrations (0.02-2.2 µL/mL). Contact activation time 5 min. Final phospholipid concentration 35 µM. Each bar represents the mean result from 3 determinations of 2-3 dilutions. For comparison, pooled mean potencies for different methods are inserted: A) Pooled mean potencies obtained with Pathromtin SL and colloidal silica containing PL-TGT and PL-PF3; n = 15. B) Pooled mean potencies obtained with SynthAFax, Rox Factor IX and PL-TGT, PL-PF3 with FXIa; n = 34.

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